

PERSPECTIVES

Nitrogen-Containing Bisphosphonates and Human $\gamma\delta$ T Cells

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Abstract

Bisphosphonates, especially nitrogen-containing bisphosphonates (N-BPs), are widely used to block bone destruction associated with bone metastasis because they are effective inhibitors of osteoclast-mediated bone resorption. In addition to their antiresorptive activity, growing preclinical evidence shows that N-BPs have direct and indirect anti-tumor activities. Some of the anti-tumor activities of N-BPs are associated with human $\gamma\delta$ T cells that are key players in the interface between innate and adaptive immunity. This review examines the molecular and cellular mechanisms through which N-BPs stimulate the expansion and anti-tumor activity of human $\gamma\delta$ T cells, suggesting a new role for N-BPs in cancer immunotherapy. *IBMS BoneKEy*. 2010 June;7(6):208-217.

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Introduction

Bisphosphonates are synthetic analogues of the naturally occurring pyrophosphate molecule that have the ability to bind to bone mineral and inhibit osteoclast-mediated bone resorption (1). Bisphosphonates may be broadly classified on the basis of whether or not they contain a nitrogen atom, with nitrogen-containing bisphosphonates (N-BPs) being more potent than non N-BPs at inhibiting osteoclast activity (1). Bisphosphonates that lack a nitrogen atom are metabolized into non-hydrolyzable pro-apoptotic ATP analogues that accumulate in the cytosol of osteoclasts (1). By contrast, N-BPs interfere with a specific enzyme in the mevalonate pathway, farnesyl pyrophosphate synthase (FPPS), thereby depleting the osteoclasts of isoprenoid lipids (2-5). More specifically, FPPS inhibition by N-BPs blocks the covalent attachment of isoprenyl chains to small GTPases (e.g., Ras, Rac, Rho, and CDC42), which is crucial for their intracellular localization and functions in osteoclasts. In addition to the effects on the function of small GTPases, the disruption of the mevalonate pathway by N-BPs results in the accumulation of

isopentenyl pyrophosphate (IPP), which is then converted (most probably via aminoacyl tRNA-synthase) to a cytotoxic adenosine triphosphate analogue (Apppl) that can directly induce osteoclast apoptosis (6,7). Thus, N-BPs may exert their pharmacological effects on osteoclasts through the formation of Apppl or via the inhibition of protein prenylation, particularly of small GTPases.

N-BPs also exert indirect and direct anticancer activities by interacting with monocytes, macrophages, endothelial and tumor cells (8). In addition, they specifically stimulate the expansion and antitumor activity of a subset of human $\gamma\delta$ T cells (referred to as $V\gamma9V\delta2$ or $V\gamma2V\delta2$ T cells), which are strongly activated by natural phosphoantigens from bacteria, parasites and eukaryotic cells (8;9). In this review, we focus on the cellular and molecular mechanisms through which N-BPs stimulate the expansion and cytotoxic activity of human $V\gamma9V\delta2$ T cells and discuss the preclinical evidence that N-BPs may have a role in cancer immunotherapy.

Activation of Human V γ 9V δ 2 T Cells by Phosphoantigens

The immune system can be divided into innate and adaptive immunity. The innate immune system serves as the first line of host defense against infection and involves non-specific guardians (*e.g.*, macrophages, dendritic cells, natural killer cells, $\gamma\delta$ T cells) (9). Then, the adaptive immune system, mediated by CD4⁺ and CD8⁺ $\alpha\beta$ T cells and B cells, provides antigen-specific lasting immunity (*e.g.*, antibody-producing and antigen-targeted cytotoxic cells) (9).

Human $\gamma\delta$ T cells are cytotoxic CD3⁺CD4⁻CD8⁻ lymphocytes that are at the interface between innate and adaptive immunity (9;10). They exhibit features of the innate immune system because, unlike $\alpha\beta$ T cells, antigen recognition by the $\gamma\delta$ T cell receptor (TCR) is not constrained by the requirement to bind major histocompatibility complex (MHC), thereby allowing $\gamma\delta$ T cells to recognize a wide range of nonpeptide phosphoantigens (9;10). Conversely, the ability of $\gamma\delta$ T cells to undergo major clonal expansion in primary infection and to mount rapid memory-type expansion upon reinfection is a feature of the adaptive immune system (10). The expression of TCR variable segments in $\gamma\delta$ T cells is associated with tissue prevalence. For example, 70% of the human $\gamma\delta$ T cells that circulate in the blood and reside in lymphoid organs express the V γ 9V δ 2 TCR, whereas $\gamma\delta$ T cells residing in the skin, gut and lungs express the V δ 1 segment and different V γ segments (9;10).

Circulating V γ 9V δ 2 T cells represent 1 to 5% of whole peripheral blood T cells and they exist only in primates and humans. They recognize non-peptide phosphorylated intermediates of bacterial isoprenoid biosynthesis (*i.e.*, (E)-4-hydroxy-3-methylbut-2-enyl pyrophosphate [HMBPP]) and phosphorylated metabolites of the mevalonate pathway in eukaryotic cells (IPP, dimethylallyl pyrophosphate [DMAPP], farnesyl pyrophosphate [FPP] and geranylgeranyl pyrophosphate [GGPP]) (9;11). Synthetic antigens such as

Phosphostim (BrHPP) are also recognized by V γ 9V δ 2 T cells (11). HMBPP is very similar in structure to IPP, but is much more potent in stimulating proliferation of V γ 9V δ 2 T cells (half-maximal concentration for stimulation of proliferation are 0.00032 and 1 μ M, respectively) (11). Other mevalonate metabolites (DMAPP, FPP, GGPP) have considerably lower potency (30- to 300-fold less than IPP) (11). The mevalonate metabolite Apppl has little stimulatory activity on V γ 9V δ 2 T cells; it could represent an inactive storage form of phosphoantigen that would require conversion to IPP to activate $\gamma\delta$ T cells (12). Thus, phosphorylated mevalonate metabolites activate V γ 9V δ 2 T cells only at high concentrations. However, certain tumors do produce, under basal conditions, elevated endogenous concentrations of IPP (*e.g.*, the B cell lymphoma cell line Daudi), which can then be sensed by V γ 9V δ 2 T cells as a tumor antigen (9;11).

The exact mechanisms through which V γ 9V δ 2 T cells become activated by IPP are still unclear, but are most likely $\gamma\delta$ TCR-mediated and require cell-cell contacts. Following activation by IPP or other phosphoantigens, V γ 9V δ 2 T cells produce pro-inflammatory chemokines (*e.g.*, MIP1 α , RANTES), interleukins (*e.g.*, GM-CSF, TGF- β), Th1 (*e.g.*, INF- γ , TNF- α) but not Th2 cytokines (*e.g.*, IL4, IL5) (13;14), and finally proliferate in the presence of IL2 (9-11). In addition, these cytokines secreted from activated $\gamma\delta$ T cells are able to regulate and stimulate other immune cells. For example, Ismaili *et al.* (15) demonstrated that $\gamma\delta$ T cell-derived TNF- α induced the maturation of dendritic cells from monocytes. Human $\gamma\delta$ T cells could also function to help initiate adaptive $\alpha\beta$ T cell responses by antigen presentation (11) and B cells for antibody production (13). Moreover, V γ 9V δ 2 T cells activated by the synthetic phosphoantigen Picostim antagonize IL2-induced expansion of Foxp3⁺ T regulatory cells (Tregs) (16). These findings (16) are of importance because Tregs play a role in mediating a balance between immunity and tolerance; they suppress the activation, proliferation

and functions of various immune cells (17). More specifically, Tregs inhibit anticancer immune responses (17) and, for example, the proliferation of human $\gamma\delta$ T cells induced by Phosphostim plus IL-2 can be suppressed by Tregs (18). It remains to be determined whether N-BPs, by promoting activation of $V\gamma9V\delta2$ T cells through IPP production, could counteract Tregs' activity, thereby facilitating cancer immunity.

Activation of Human $V\gamma9V\delta2$ T Cells by N-BPs

Evidence for the stimulation of $V\gamma9V\delta2$ T cells by N-BPs was first found when increased numbers of $\gamma\delta$ T cells were observed in patients who had flu-like acute-phase reactions after their first intravenous infusion of pamidronate (19). N-BPs might induce or activate $V\gamma9V\delta2$ T cells either by mimicking phosphoantigens and/or by increasing phosphoantigen levels. For example, N-BPs (pamidronate, risedronate, zoledronate), in the presence of low doses of IL-2, can activate and stimulate the proliferation of human $V\gamma9V\delta2$ T cells *in vitro* and *in vivo* (11;20). Furthermore, zoledronate induces functional changes in $V\gamma9V\delta2$ T cell subsets (8;21). *In vivo*, it promotes the differentiation of $V\gamma9V\delta2$ T cells toward $CD45RA^+CD27^-$ $\gamma\delta$ T cells, which produce interferon- γ and exert cytotoxicity, while decreasing $CD45RA^+CD27^+$ naive and $CD45RA^+CD27^+$ memory $\gamma\delta$ T cells (8;21). This effect is specific to $V\gamma9V\delta2$ T cells. Neither human $\gamma\delta$ T cells expressing the $V\gamma9V\delta1$ TCR, nor human $\alpha\beta$ T cells, monocytes, NK or B cells are responsive to N-BPs (11;21). These results suggested a direct binding of N-BPs to the TCR expressed by human $V\gamma9V\delta2$ T cells. However, it has since become clear that the activation of human $\gamma\delta$ T cells by N-BPs requires both antigen-presenting cells (monocytes, dendritic cells) and the inhibition of the mevalonate pathway (11;21). For instance, zoledronic acid promotes the immunostimulatory properties of human dendritic cells by enhancing their ability to activate $\gamma\delta$ T cells (22). Similarly, pamidronate-treated, but not untreated,

THP-1 monocytic cells are capable of activating purified $\gamma\delta$ T cells to produce interferon- γ (23). In addition, pamidronate-treated THP-1 cells activate TCR-defective Jurkat cells only when these cells are stably transfected to express the $V\gamma9V\delta2$ TCR (23;24). These results (23;24) suggest therefore that N-BP-treated antigen-presenting cells (as exemplified by THP-1 cells) could activate $\gamma\delta$ T cells in a TCR-dependent manner. Because N-BPs specifically inhibit the IPP-consuming enzyme FPPS, IPP could then be directly recognized by $\gamma\delta$ T cells. For example, zoledronic acid induces the accumulation of IPP in monocytes from human peripheral blood mononuclear cells (PBMCs) which, in turn, activates the expansion of $\gamma\delta$ T cells in a cell contact-dependent manner (25). In addition, the expansion of $\gamma\delta$ T cells from PBMCs treated with a N-BP is prevented by statins, which inhibit HMG-CoA reductase upstream of FPP synthase and prevent the synthesis of IPP (26). Thus, the internalization of N-BPs by monocytes and dendritic cells, which are highly endocytic cells, leads to the inhibition of the mevalonate pathway and subsequent intracellular accumulation of IPP which, in turn, activates $\gamma\delta$ T cells. Mechanisms through which mevalonate metabolites (such as IPP) are cell-surface exposed and recognized by the $V\gamma9V\delta2$ TCR are, however, still unknown. Attempts to co-crystallize IPP (or HMBPP) with the $V\gamma9V\delta2$ TCR have not succeeded (27). However, it has been shown that there exists a protein-associated membrane component on the antigen-presenting cell surface that presents HMBPP to the $V\gamma9V\delta2$ TCR for immune recognition (28). It is therefore most conceivable that IPP may also be complexed with a cell surface antigen-presenting molecule in order to be recognized by the $V\gamma9V\delta2$ TCR.

Preclinical Evidence for a Role of N-BPs in Promoting Cancer Immunotherapy

Most of the human tumor cell lines treated with a N-BP can efficiently activate human $\gamma\delta$ T cells to proliferate and lyse tumor cells in a $\gamma\delta$ TCR-dependent manner (24). In

contrast, tumor cell lines of nonhuman origins treated with a N-BP fail to activate human V γ 9V δ 2 T cells, indicating species-specific cell-cell interactions (29). These findings (29) are in line with the observation that nonhuman antigen-presenting cells do not have a protein-associated membrane component that presents HMBPP for recognition by the V γ 9V δ 2 TCR (28).

N-BPs induce intracellular accumulation of IPP/Apppl in a wide variety of human tumor cell lines (30;31) and these mevalonate metabolites could be sensed by V γ 9V δ 2 T cells as tumor phosphoantigens. This contention is supported by the observation that the silencing of FPPS protein expression by a shRNA in Raji and HepG2 tumor cells converts these cells into V γ 9V δ 2 T cell activators (32), presumably because of the higher intracellular IPP levels. In addition, mevastatin and lovastatin (HMG-CoA reductase inhibitors that prevent the synthesis of IPP) completely abolish V γ 9V δ 2 T cell activation induced by zoledronate- or pamidronate-treated tumor cells (Daudi lymphoma, K562 leukemia, KMM1 myeloma and colon carcinoma cell lines) (12;33-35). Thus, tumor cells that are treated with N-BPs overproduce mevalonate metabolites (such as IPP) that are somehow sensed by V γ 9V δ 2 T cells as tumor antigens, causing their activation and then the efficient killing of bisphosphonate-treated tumor cells. As aforementioned for HMBPP (28), it is most unlikely that IPP directly binds to the V γ 9V δ 2 TCR. Instead, IPP (or Apppl) may be complexed with an antigen-presenting molecule in order to be recognized by the V γ 9V δ 2 TCR (11). In this respect, the V γ 9V δ 2 TCR binds to a complex formed between apolipoprotein A1 and F1-ATP synthase (AS), which is a mitochondrial enzyme that is translocated to the cell surface of tumor cells (36). Interestingly, Apppl inhibits a mitochondrial ADP/ATP translocase (6), suggesting that Apppl might also bind to AS in order to be recognized by the V γ 9V δ 2 TCR (36). It has also been proposed that Apppl could represent an inactive storage form of phosphoantigen that, when exposed at the tumor cell surface, would require hydrolysis by some

ecto-nucleotide pyrophosphatases for conversion into IPP (12). Thus, upon treatment of tumor cells with a N-BP, Apppl could be translocated to the tumor cell surface as a complex with a putative antigen-presenting molecule and then processed into IPP for its subsequent recognition by the V γ 9V δ 2 TCR. Although highly speculative, this hypothesis clearly warrants further investigation.

Other cell surface receptors have been involved in mediating cell-cell interactions between V γ 9V δ 2 T cells and N-BP-treated tumor cells (Table 1). For example, LFA1 expressed on $\gamma\delta$ T cells mediates a stable interaction with pamidronate- or zoledronate-treated tumor cells expressing ICAM-1 (29;37). However, relatively limited types of tumor cells express these molecules. V γ 9V δ 2 T cells also express the NKG2D receptor, a type II C-lectin-like protein that is expressed by NK cells. NKG2D interacts with MHC class I-related chains A/B (MICA/MICB) and the UL-16 binding proteins 1 to 4 (ULBP1-4) that are frequently expressed by tumor cells (colon and renal carcinomas, myelomas, lymphomas) and thus can contribute to the efficient killing of N-BP-treated tumor cells by activated V γ 9V δ 2 T cells (8;11). The engagement of CD6 on human $\gamma\delta$ T cells by CD166 on human tumor cells also seems to play an important role in bisphosphonate-mediated $\gamma\delta$ T-cell activation (38). Expression of CD166 has been described in malignant melanoma and various carcinomas (breast, prostate, lung, colon, and bladder) (8). Its *de novo* expression in CD166-negative K562 leukemia cells markedly enhances the activation of $\gamma\delta$ T cells following pamidronate treatment of CD166-expressing K562 cells (38). Conversely, the silencing of CD166 in LK-2 lung carcinoma cells treated with pamidronate decreases $\gamma\delta$ T cell activation (38). Thus, upon treatment of tumor cells with N-BPs, CD6/CD166 and NKG2D/MICA/B interactions could provide costimulatory signals for TCR-mediated $\gamma\delta$ T cell activation (Table 1). In addition, it has been reported that only IPP-activated $\gamma\delta$ T cells expressing CD56 can efficiently kill

Table 1. Contribution of cell surface receptors in activated human V γ 9V δ 2 T cells to recognition of bisphosphonate-treated tumor cells.

$\gamma\delta$ T cell surface receptor	Tumor cell surface ligand(s)	Ref.
γ 9 δ 2 TCR	F1-ATP synthase, apolipoprotein A1	(36)
NKG2D	MICA, MICB, ULBP 1-4	(8;11)
LFA1	ICAM-1	(29;37)
CD6	CD166	(38)
CD56	unidentified	(39)

tumor cells because they secrete increased amounts of cytolytic granules (granzyme B, perforin) when compared to CD56-negative $\gamma\delta$ T cells (39) (Table 1).

In vivo experimental studies demonstrate that zoledronate significantly enhances the anti-tumor activity of purified human V γ 9V δ 2 T cells, which have been transferred into immunodeficient mice xenografted with SBC-5 small cell lung carcinoma cells (40), UM-UC-3 bladder cancer cells (41) or MM1 chronic myelogenous leukemic cells (42). Similarly, upon transfer into SCID mice, purified human V γ 9V δ 2 T cells given together with alendronate plus IL-2 significantly prolong the survival of animals bearing MeWo melanoma cells or PancTu1 pancreatic carcinoma cells (43). Importantly, V γ 9V δ 2 T cells can be expanded from PBMCs of patients with cancer (chronic myeloid leukemia, multiple myeloma, breast and prostate carcinomas), following treatment with zoledronate (or Phosphostim) plus IL-2 (34;42;44-47). These cells exhibit potent anti-tumor activity *in vitro* against human cancer cell lines (33;34;44-46). In addition, V γ 9V δ 2 T cells expanded from PBMCs of patients with chronic myeloid leukemia also exhibit antitumor activity *in vitro* against autologous or allogeneic, zoledronate-treated leukemia cells isolated from patients with chronic myeloid leukemia (42).

Furthermore, *in vitro* experiments showed that zoledronate enhances the chemotherapy-induced sensitization of

different tumor cell lines to V γ 9V δ 2 T cell cytotoxicity (44). It also sensitizes human colon cancer stem cells (which are more resistant to chemotherapy) and imatinib-resistant chronic myeloid leukemic cell lines to V γ 9V δ 2 T cell-mediated killing *in vitro* (35;42).

Concluding Comments and Future Directions

Studies on the effect of N-BPs on human $\gamma\delta$ T cells have revealed a previously unappreciated approach to exploit the antitumor potential of bisphosphonates. These studies have shown that by blocking FPPS activity with N-BPs it is possible to cause intracellular accumulation of IPP/Apppl both in tumor cells and antigen-presenting cells (monocytes, dendritic cells), thereby leading to V γ 9V δ 2 T cell antitumor activity. Future work should aim at characterizing molecular mechanisms responsible for the cell surface exposure and recognition of IPP/Apppl by the V γ 9V δ 2 TCR. However, these preclinical studies already suggest that N-BPs could be used as a promising immunotherapeutic approach for V γ 9V δ 2 T cell activation. Two strategies for the potential usage of $\gamma\delta$ T cells in cancer immunotherapy are currently under clinical investigation: (1) the adoptive transfer of *ex vivo* expanded autologous V γ 9V δ 2 T cells and (2) the *in vivo* activation of V γ 9V δ 2 T cells. Abe *et al.* (48) conducted a phase I clinical trial in multiple myeloma patients using adoptive transfer of autologous

Table 2. Therapeutic activation of V γ 9V δ 2 T cells by bisphosphonates in early clinical trials

Cancer type	Treatment	Observations	Ref.
Multiple myeloma	zoledronate + IL-2	Study in 52 patients. Induction of $\gamma\delta$ T cell effector functions in half of the patients.	(34)
Low-grade non-Hodgkin lymphoma	pamidronate + IL-2	Antitumor activity was noted when patients who responded to pamidronate <i>in vitro</i> were treated (50% of patients). In this respect, a significant <i>in vivo</i> activation of $\gamma\delta$ T cells and objective clinical responses in 5 out of 9 patients were reported.	(50)
Breast carcinoma	zoledronate	Study in 23 patients. A single dose of zoledronate induces a long-lasting activation of $\gamma\delta$ T cells.	(49)
Prostate carcinoma	zoledronate + IL-2	Study in 18 patients. Induction of $\gamma\delta$ T cell effector functions and improvement of clinical responses.	(47)

V γ 9V δ 2 T cells generated from PBMCs after *ex vivo* incubation with zoledronate plus IL-2. The authors reported an increased number of CD45RA-CD27- effector memory cells in the blood and bone marrow from these patients, 4 weeks after initiation of treatment (48). Regarding the second immunotherapeutic strategy that consists of the *in vivo* activation of $\gamma\delta$ T cells, a study showed that treatment of breast cancer patients with zoledronate without IL-2 led to an increase in the percentage of effector V γ 9V δ 2 T cells in blood (49). Moreover, early clinical trials with N-BPs (pamidronate, zoledronate) plus IL-2 have been performed in patients with multiple myeloma (34), prostate carcinoma (47) and lymphoma (50), and data showed a significant expansion of V γ 9V δ 2 T cells in several cancer patients and even good clinical responses in some prostate cancer patients (Table 2). These results suggest, therefore, that there is an interest in (and rationale for) using N-BPs for *ex vivo* or *in vivo* activation of V γ 9V δ 2 T cells. However, about half of the patients enrolled in these clinical trials failed to expand their V γ 9V δ 2 T cells after treatment with a N-BP plus IL-2 (Table 2). This kind of $\gamma\delta$ T cell anergy is frequently observed in cancer patients (34;50). It might be related to the negative effect of Tregs on V γ 9V δ 2 T

cell expansion (9;17;18). Alternatively, there might be a progressive deterioration of V γ 9V δ 2 T cell immunity along with disease progression. Future challenges will be, therefore, to optimize immunotherapeutic protocols that both target V γ 9V δ 2 T cell expansion and overcome $\gamma\delta$ T cell anergy. Combination approaches that associate V γ 9V δ 2 T cell-based immunotherapy with chemotherapy might be another promising therapeutic strategy to sensitize tumor cells to V γ 9V δ 2 T cell cytotoxicity.

Conflict of Interest: Dr. Clézardin reports that he has served on advisory boards for Novartis, and has received lecture fees and research support from Novartis and Procter & Gamble Pharmaceuticals. Dr. Benzaid: none reported.

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References

1. Fleisch H. Development of bisphosphonates. *Breast Cancer Res.* 2002;4(1):30-4.
2. Luckman SP, Coxon FP, Ebetino FH, Russell RG, Rogers MJ. Heterocycle-containing bisphosphonates cause apoptosis and inhibit bone resorption by preventing protein prenylation: evidence

- from structure-activity relationships in J774 macrophages. *J Bone Miner Res.* 1998 Nov;13(11):1668-78.
3. Luckman SP, Hughes DE, Coxon FP, Graham R, Russell G, Rogers MJ. Nitrogen-containing bisphosphonates inhibit the mevalonate pathway and prevent post-translational prenylation of GTP-binding proteins, including Ras. *J Bone Miner Res.* 1998 Apr;13(4):581-9.
 4. Kavanagh KL, Guo K, Dunford JE, Wu X, Knapp S, Ebetino FH, Rogers MJ, Russell RG, Oppermann U. The molecular mechanism of nitrogen-containing bisphosphonates as antiosteoporosis drugs. *Proc Natl Acad Sci U S A.* 2006 May 16;103(20):7829-34.
 5. Rondeau JM, Bitsch F, Bourgier E, Geiser M, Hemmig R, Kroemer M, Lehmann S, Ramage P, Rieffel S, Strauss A, Green JR, Jahnke W. Structural basis for the exceptional in vivo efficacy of bisphosphonate drugs. *ChemMedChem.* 2006 Feb;1(2):267-73.
 6. Mönkkönen H, Auriola S, Lehenkari P, Kellinsalmi M, Hassinen IE, Vepsäläinen J, Mönkkönen J. A new endogenous ATP analog (Apppl) inhibits the mitochondrial adenine nucleotide translocase (ANT) and is responsible for the apoptosis induced by nitrogen-containing bisphosphonates. *Br J Pharmacol.* 2006 Feb;147(4):437-45.
 7. Rääkkönen J, Crockett JC, Rogers MJ, Mönkkönen H, Auriola S, Mönkkönen J. Zoledronic acid induces formation of a pro-apoptotic ATP analogue and isopentenyl pyrophosphate in osteoclasts in vivo and in MCF-7 cells in vitro. *Br J Pharmacol.* 2009 Jun;157(3):427-35.
 8. Stresing V, Daubiné F, Benzaid I, Mönkkönen H, Clézardin P. Bisphosphonates in cancer therapy. *Cancer Lett.* 2007 Nov 8;257(1):16-35.
 9. Kabelitz D, Wesch D, He W. Perspectives of $\gamma\delta$ T cells in tumor immunology. *Cancer Res.* 2007 Jan 1;67(1):5-8.
 10. Carding SR, Egan PJ. Gammadelta T cells: functional plasticity and heterogeneity. *Nat Rev Immunol.* 2002 May;2(5):336-45.
 11. Morita CT, Jin C, Sarikonda G, Wang H. Nonpeptide antigens, presentation mechanisms, and immunological memory of human Vgamma2Vdelta2 T cells: discriminating friend from foe through the recognition of prenyl pyrophosphate antigens. *Immunol Rev.* 2007 Feb;215:59-76.
 12. Vantourout P, Mookerjee-Basu J, Rolland C, Pont F, Martin H, Davrinche C, Martinez LO, Perret B, Collet X, Périgaud C, Peyrottes S, Champagne E. Specific requirements for Vgamma9Vdelta2 T cell stimulation by a natural adenylated phosphoantigen. *J Immunol.* 2009 Sep 15;183(6):3848-57.
 13. Caccamo N, Battistini L, Bonneville M, Poccia F, Fournié JJ, Meraviglia S, Borsellino G, Kroczeck RA, La Mendola C, Scotet E, Dieli F, Salerno A. CXCR5 identifies a subset of Vgamma9Vdelta2 T cells which secrete IL-4 and IL-10 and help B cells for antibody production. *J Immunol.* 2006 Oct 15;177(8):5290-5.
 14. Wesch D, Glatzel A, Kabelitz D. Differentiation of resting human peripheral blood gamma delta T cells toward Th1- or Th2-phenotype. *Cell Immunol.* 2001 Sep 15;212(2):110-7.
 15. Ismaili J, Olislagers V, Poupot R, Fournié JJ, Goldman M. Human gamma delta T cells induce dendritic cell maturation. *Clin Immunol.* 2002 Jun;103(3 Pt 1):296-302.
 16. Gong G, Shao L, Wang Y, Chen CY, Huang D, Yao S, Zhan X, Sicard H, Wang R, Chen ZW. Phosphoantigen-activated V gamma 2V delta 2 T cells antagonize IL-2-induced

- CD4+CD25+Foxp3+ T regulatory cells in mycobacterial infection. *Blood*. 2009 Jan 22;113(4):837-45.
17. Mouggiakakos D, Choudhury A, Lladser A, Kiessling R, Johansson CC. Regulatory T cells in cancer. *Adv Cancer Res*. 2010;107:57-117.
 18. Kunzmann V, Kimmel B, Herrmann T, Einsele H, Wilhem M. Inhibition of phosphoantigen-mediated gammadelta T-cell proliferation by CD4+CD25+FoxP3+ regulatory T cells. *Immunology*. 2009 Feb;126(2):256-67.
 19. Kunzmann V, Bauer E, Wilhelm M. Gamma/delta T-cell stimulation by pamidronate. *N Engl J Med*. 1999 Mar 4;340(9):737-8.
 20. Casetti R, Perretta G, Taglioni A, Mattei M, Colizzi V, Dieli F, D'Offizi G, Malkovsky M, Poccia F. Drug-induced expansion and differentiation of V gamma delta 2 T cells in vivo: the role of exogenous IL-2. *J Immunol*. 2005 Aug 1;175(3):1593-8.
 21. Caccamo N, Meraviglia S, Scarpa F, La Mendola C, Santini D, Bonanno CT, Misiano G, Dieli F, Salerno A. Aminobisphosphonate-activated gamma delta T cells in immunotherapy of cancer: doubts no more. *Expert Opin Biol Ther*. 2008 Jul;8(7):875-83.
 22. Fiore F, Castella B, Nuschak B, Bertieri R, Mariani S, Bruno B, Pantaleoni F, Foglietta M, Boccadoro M, Massaia M. Enhanced ability of dendritic cells to stimulate innate and adaptive immunity on short-term incubation with zoledronic acid. *Blood*. 2007 Aug 1;110(3):921-7.
 23. Miyagawa F, Tanaka Y, Yamashita S, Minato N. Essential requirement of antigen presentation by monocyte lineage cells for the activation of primary human gamma delta T cells by aminobisphosphonate antigen. *J Immunol*. 2001 May 1;166(9):5508-14.
 24. Tanaka Y. Human gammadelta T cells and tumor immunotherapy. *J Clin Exp Hematopathol*. 2006;46:11-23.
 25. Roelofs AJ, Jauhainen M, Mönkkönen H, Rogers MJ, Mönkkönen J, Thompson K. Peripheral blood monocytes are responsible for gammadelta T cell activation induced by zoledronic acid through accumulation of IPP/DMAPP. *Br J Haematol*. 2009 Jan;144(2):245-50.
 26. Thompson K, Rogers MJ. Statins prevent bisphosphonate-induced gamma,delta-T-cell proliferation and activation in vitro. *J Bone Miner Res*. 2004 Feb;19(2):278-88.
 27. Allison TJ, Winter CC, Fournié JJ, Bonneville M, Garboczi DN. Structure of a human gammadelta T-cell antigen receptor. *Nature*. 2001 Jun 14;411(6839):820-4.
 28. Wei H, Huang D, Lai X, Chen M, Zhong W, Wang R, Chen ZW. Definition of APC presentation of phosphoantigen (E)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate to Vgamma2Vdelta 2 TCR. *J Immunol*. 2008 Oct 1;181(7):4798-806.
 29. Kato Y, Tanaka Y, Tanaka H, Yamashita S, Minato N. Requirement of species-specific interactions for the activation of human gamma delta T cells by pamidronate. *J Immunol*. 2003 Apr 1;170(7):3608-13.
 30. Mönkkönen H, Kuokkanen J, Holen I, Evans A, Lefley DV, Jauhainen M, Auriola S, Mönkkönen J. Bisphosphonate-induced ATP analog formation and its effect on inhibition of cancer cell growth. *Anticancer Drugs*. 2008 Apr;19(4):391-9.
 31. Mitrofan LM, Pelkonen J, Mönkkönen J. The level of ATP analog and isopentenyl pyrophosphate correlates with zoledronic acid-induced apoptosis in cancer cells in vitro. *Bone*. 2009 Dec;45(6):1153-60.

32. Li J, Herold MJ, Kimmel B, Müller I, Rincon-Orozco B, Kunzmann V, Herrmann T. Reduced expression of the mevalonate pathway enzyme farnesyl pyrophosphate synthase unveils recognition of tumor cells by Vgamma9Vdelta2 T cells. *J Immunol*. 2009 Jun 15;182(12):8118-24.
33. Gober HJ, Kistowska M, Angman L, Jenö P, Mori L, De Libero G. Human T cell receptor gammadelta cells recognize endogenous mevalonate metabolites in tumor cells. *J Exp Med*. 2003 Jan 20;197(2):163-8.
34. Mariani S, Muraro M, Pantaleoni F, Fiore F, Nuschak B, Peola S, Foglietta M, Palumbo A, Coscia M, Castella B, Bruno B, Bertieri R, Boano L, Boccadoro M, Massaia M. Effector gammadelta T cells and tumor cells as immune targets of zoledronic acid in multiple myeloma. *Leukemia*. 2005 Apr;19(4):664-70.
35. Todaro M, D'Asaro M, Caccamo N, Iovino F, Francipane MG, Meraviglia S, Orlando V, La Mendola C, Gulotta G, Salerno A, Dieli F, Stassi G. Efficient killing of human colon cancer stem cells by gammadelta T lymphocytes. *J Immunol*. 2009 Jun 1;182(11):7287-96.
36. Bonneville M, Scotet E. Human Vgamma9Vdelta2 T cells: promising new leads for immunotherapy of infections and tumors. *Curr Opin Immunol*. 2006 Oct;18(5):539-46.
37. Uchida R, Ashihara E, Sato K, Kimura S, Kuroda J, Takeuchi M, Kawata E, Taniguchi K, Okamoto M, Shimura K, Kiyono Y, Shimazaki C, Taniwaki M, Maekawa T. Gamma delta T cells kill myeloma cells by sensing mevalonate metabolites and ICAM-1 molecules on cell surface. *Biochem Biophys Res Commun*. 2007 Mar 9;354(2):613-8.
38. Kato Y, Tanaka Y, Hayashi M, Okawa K, Minato N. Involvement of CD166 in the activation of human gamma delta T cells by tumor cells sensitized with nonpeptide antigens. *J Immunol*. 2006 Jul 15;177(2):877-84.
39. Alexander AA, Maniar A, Cummings JS, Hebbeler AM, Schulze DH, Gastman BR, Pauza CD, Strome SE, Chapoval AI. Isopentenyl pyrophosphate-activated CD56+ gammadelta T lymphocytes display potent antitumor activity toward human squamous cell carcinoma. *Clin Cancer Res*. 2008 Jul 1;14(13):4232-40.
40. Sato K, Kimura S, Segawa H, Yokota A, Matsumoto S, Kuroda J, Nogawa M, Yuasa T, Kiyono Y, Wada H, Maekawa T. Cytotoxic effects of gammadelta T cells expanded ex vivo by a third generation bisphosphonate for cancer immunotherapy. *Int J Cancer*. 2005 Aug 10;116(1):94-9.
41. Yuasa T, Sato K, Ashihara E, Takeuchi M, Maita S, Tsuchiya N, Habuchi T, Maekawa T, Kimura S. Intravesical administration of gammadelta T cells successfully prevents the growth of bladder cancer in the murine model. *Cancer Immunol Immunother*. 2009 Apr;58(4):493-502.
42. D'Asaro M, La Mendola C, Di Liberto D, Orlando V, Todaro M, Spina M, Guggino G, Meraviglia S, Caccamo N, Messina A, Salerno A, Di Raimondo F, Vigneri P, Stassi G, Fournié JJ, Dieli F. V gamma 9V delta 2 T lymphocytes efficiently recognize and kill zoledronate-sensitized, imatinib-sensitive, and imatinib-resistant chronic myelogenous leukemia cells. *J Immunol*. 2010 Mar 15;184(6):3260-8.
43. Kabelitz D, Wesch D, Pitters E, Zöller M. Characterization of tumor reactivity of human V gamma 9V delta 2 gamma delta T cells in vitro and in SCID mice in vivo. *J Immunol*. 2004 Dec 1;173(11):6767-76.
44. Mattarollo SR, Kenna T, Nieda M, Nicol AJ. Chemotherapy and zoledronate sensitize solid tumour cells to Vgamma9Vdelta2 T cell cytotoxicity.

- Cancer Immunol Immunother.* 2007 Aug;56(8):1285-97.
45. Kondo M, Sakuta K, Noguchi A, Ariyoshi N, Sato K, Sato S, Sato K, Hosoi A, Nakajima J, Yoshida Y, Shiraishi K, Nakagawa K, Kakimi K. Zoledronate facilitates large-scale ex vivo expansion of functional gammadelta T cells from cancer patients for use in adoptive immunotherapy. *Cytotherapy.* 2008;10(8):842-56.
46. Burjanadzé M, Condomines M, Reme T, Quittet P, Latry P, Lugagne C, Romagne F, Morel Y, Rossi JF, Klein B, Lu ZY. In vitro expansion of gamma delta T cells with anti-myeloma cell activity by Phosphostim and IL-2 in patients with multiple myeloma. *Br J Haematol.* 2007 Oct;139(2):206-16.
47. Dieli F, Vermijlen D, Fulfarò F, Caccamo N, Meraviglia S, Cicero G, Roberts A, Buccheri S, D'Asaro M, Gebbia N, Salerno A, Eberl M, Hayday AC. Targeting human gammadelta T cells with zoledronate and interleukin-2 for immunotherapy of hormone-refractory prostate cancer. *Cancer Res.* 2007 Aug 1;67(15):7450-7.
48. Abe Y, Muto M, Nieda M, Nakagawa Y, Nicol A, Kaneko T, Goto S, Yokokawa K, Suzuki K. Clinical and immunological evaluation of zoledronate-activated Vgamma9gammadelta T-cell-based immunotherapy for patients with multiple myeloma. *Exp Hematol.* 2009 Aug;37(8):956-68.
49. Santini D, Martini F, Fratto ME, Galluzzo S, Vincenzi B, Agrati C, Turchi F, Piacentini P, Rocci L, Manavalan JS, Tonini G, Poccia F. In vivo effects of zoledronic acid on peripheral gammadelta T lymphocytes in early breast cancer patients. *Cancer Immunol Immunother.* 2009 Jan;58(1):31-8.
50. Wilhelm M, Kunzmann V, Eckstein S, Reimer P, Weissinger F, Ruediger T, Tony HP. Gammadelta T cells for immune therapy of patients with lymphoid malignancies. *Blood.* 2003 Jul 1;102(1):200-6.