

## REVIEW

# Genetic aspects of skeletal muscle strength and mass with relevance to sarcopenia

Stephen M Roth

Department of Kinesiology, School of Public Health, University of Maryland, College Park, MD, USA.

Skeletal muscle is a highly heritable quantitative trait, with heritability estimates ranging 30–85% for muscle strength and 50–80% for lean mass. That strong genetic contribution indicates the possibility of using genetic information to individualize treatments for sarcopenia or even aid in prevention strategies through the use of genetic screening prior to the functional limitations. Though these possibilities provide the rationale for genetic studies of skeletal muscle traits, few genes have been identified that appear to contribute to variation in either skeletal muscle strength or mass phenotypes, and sarcopenia *per se* is remarkably understudied as a trait in this regard. This review examines the heritability of skeletal muscle traits, findings of linkage and genome-wide association analyses and impact of specific genes and gene-sequence variants on these traits as relevant to sarcopenia. Despite considerable work in the area, the genetic underpinnings of skeletal muscle traits remain largely unknown and the genetic aspects of sarcopenia are even less clear. Large-scale longitudinal clinical studies relying on advanced genome-wide association and other techniques are needed to provide further insights into the genes and gene variants that contribute to skeletal muscle strength and mass, and ultimately to susceptibility to sarcopenia.

*BoneKEy Reports* 1, Article number: 58 (2012) | doi:10.1038/bonekey.2012.58

### Introduction

Aging is associated with a decline in skeletal muscle strength, mass, power and physical functioning known as sarcopenia.<sup>1</sup> These losses have important health consequences,<sup>2–5</sup> including an increased risk of falls, hip fractures and functional decline.<sup>4–6</sup> Muscle strength is independently associated with functional ability in the elderly<sup>5,7–9</sup> and may explain up to 25% of the variance in overall functional ability.<sup>10</sup> Sarcopenia is also related to a reduction in the performance of activities of daily living,<sup>11</sup> which may lead to further declines in muscle mass and strength and greater reductions in the performance of those activities. The overall effect of this cycle can be a marked loss of function, predisposing older individuals to falls, injuries and disability.<sup>12</sup>

Although the loss of muscle mass is associated with the decline in strength with advancing age, the strength decline is much more rapid than the accompanying loss of muscle mass, indicating a decline in muscle quality.<sup>13</sup> In fact, the loss of muscle strength is a stronger predictor of mortality in the elderly than the loss of muscle mass.<sup>12,14–16</sup> The relationships of muscle mass and strength to mortality appear to lie in the higher functional capacity associated with greater muscle strength regardless of mass, resulting in an inverse association with functional limitations and disability. The consequences of sarcopenia-related disability are significant both in terms of quality

of life and health-care costs related to sarcopenia, estimated to be \$18.5 billion dollars in the United States for adults  $\geq 60$  years for the year 2000.<sup>17</sup>

Though the losses of muscle mass and strength begin on average between 40 and 50 years of age, losses for any particular individual are difficult to predict. Sarcopenia has been reported in community-dwelling men and women  $< 50$  years,<sup>4,5,18,19</sup> and sarcopenia associated with compromised physical functioning occurs in nearly one in ten women aged 34–58 years,<sup>20</sup> providing further support for the variable onset of muscle strength losses and an indication of susceptibility to sarcopenia in some individuals. Various research groups are currently exploring the possibility that a portion of this inter-individual variability and susceptibility to early muscle losses is due to genetic factors, which is the focus of the present review. The importance of physical activity and resistance training particularly in slowing the losses of muscle mass and strength is clear<sup>21,22</sup> and genetic factors have been found important in this context,<sup>23,24</sup> but a discussion of that related literature is beyond the scope of the present review. Similarly, the present review focuses on human studies; readers are pointed to a more comprehensive review published recently<sup>25</sup> that discusses key findings from animal models that speak to the genetic aspects of skeletal muscle traits.

Correspondence: Dr SM Roth, Department of Kinesiology, School of Public Health, University of Maryland, 2134C SPH Building, College Park, MD, USA.  
E-mail: sroth1@umd.edu

Received 21 December 2011; accepted 2 March 2012; published online 4 April 2012

## Heritability of Skeletal Muscle Traits

Variation in skeletal muscle traits among individuals can be attributed to genetic factors, environmental factors or some interaction of both. Though the influence of environmental factors, such as physical activity and diet, have been broadly investigated, only recently studies have begun to identify the specific genetic influences on skeletal muscle traits that may explain the inter-individual trait variability. Initial studies in this regard have focused on establishing the heritability of muscle-related traits. For example, the heritability of grip strength was estimated between 30 and 50% in several early studies.<sup>26–28</sup> In older twins, genetic factors accounted for 65% of the variance in grip strength even after adjusting for body weight, height and age.<sup>29</sup> More recently, twin studies have revealed heritability values for muscle strength phenotypes ranging 30–85% depending on the conditions of the strength measure (for example, limb, contraction angle, velocity and type).<sup>23,29–33</sup>

With regard to skeletal muscle mass, the first direct study of lean body mass was performed by Bouchard *et al.*<sup>34</sup> who reported 80% heritability of lean body mass by hydrodensitometry in twin pairs. Similarly, high heritability values have been reported by many other groups using more sophisticated measurement techniques.<sup>30,35–38</sup> Across these studies, heritability estimates >50% are not uncommon for muscle mass measurements.

Perhaps the most relevant for this review are the studies that have examined heritability within older subjects. Several reports have demonstrated significant heritability values for muscle strength in older individuals.<sup>29,39–43</sup> Frederiksen and colleagues<sup>39</sup> reported heritability of grip strength at 50% in individuals from 46 to 96 years. Even the change in muscle strength with advancing age has been found to be heritable,<sup>44,45</sup> though some studies indicate that the contribution of environmental factors increases at older ages.<sup>42,46</sup> Overall, genetic variation explains a significant fraction of the inter-individual variability in skeletal muscle phenotypes, including muscle traits in older individuals. Despite strong evidence for a heritable component to muscle phenotypes, the genetic underpinnings of this heritability is only beginning to be known.

## Linkage-Analysis and Genome-Wide Association Studies (GWASs) and Skeletal Muscle Traits

After the heritability of a trait is established, a common next step was to perform linkage-analysis studies in families, which sought to identify chromosome locations that harbor genes and gene variants that contribute to trait variation. Several such studies have been done for various measures of lean mass or muscle strength phenotypes and chromosome regions, and in some cases, specific gene loci have been identified as candidate regions.<sup>47–51</sup> Only one linkage study has targeted older individuals in particular. Tiainen *et al.*<sup>42</sup> examined 397 microsatellite markers in 94 female dizygotic twin pairs aged 66–75 years from the Finnish Twin Study on Aging. Significant linkages were reported for knee extensor isometric strength, leg extensor power and calf muscle cross-sectional area.

A few detailed linkage studies have been performed, isolating a small number of chromosome regions in order to better identify potential candidate genes.<sup>52–54</sup> Huygens and colleagues<sup>53</sup> performed a gene-targeted linkage analysis in the myostatin pathway (across 10 genes) in a young male cohort for various

measures of muscle mass and strength. Significant linkage was reported for knee extension and flexion peak torque measures in the *MSTN* (myostatin), *CDKN1A* and *MYOD1* genes. The same group then performed an expanded multi-point linkage analysis in 367 young Caucasian male siblings from 145 families with nine genes involved in the myostatin signaling pathway and various measures of muscle strength.<sup>54</sup> Significant linkages were reported on four chromosomal regions with knee muscle strength measures. Most recently, Windelinckx and colleagues<sup>55</sup> performed a focused fine mapping of chromosome 12q12–14 and identified *ACVR1B* as a candidate gene for muscle strength.

Linkage analysis studies have given way recently to GWAS that can be used to identify specific gene regions in unrelated individuals by use of high-density polymorphism microarrays. In the first such GWAS for lean mass, Liu and colleagues<sup>56</sup> examined 379 319 polymorphisms in nearly 1000 unrelated US whites and identified two polymorphisms both located in the thyrotropin-releasing hormone receptor (*TRHR*) gene as statistically significant. Importantly, these associations were confirmed in three replication cohorts consisting of over 6000 total white and Chinese subjects. Sun *et al.*<sup>57</sup> recently identified several novel candidate polymorphisms underlying both appendicular lean mass and femoral neck geometric properties in a large GWAS in Chinese adults followed by independent replication in white subjects. Most recently, Hai and colleagues<sup>58</sup> performed a copy-number variation GWAS in this same Chinese cohort and identified the *Gremlin1* gene as significantly associated with lean mass. These studies thus contribute to the identification of specific genes and gene variants with clinically relevant influences on skeletal muscle traits important to physical function, though they are preliminary in nature and require significant replication and follow-up work to confirm their observations.

## Genetic Variation and Skeletal Muscle Strength

Beyond the heritability and linkage or GWAS investigations, ultimately researchers are interested in identifying the specific genes and gene variants contributing to the genetic influence underlying these skeletal muscle traits. This section reviews specific genes that have been examined in multiple studies in relation to skeletal muscle strength measurements, focusing on genes associated with baseline strength values. Not all such genes are reviewed here, but the selected genes represent either those most studied or those with significant findings; a more comprehensive listing of genes was recently published.<sup>25</sup> The identification of genetic factors important to skeletal muscle strength is remarkably difficult given that multiple strength variables are commonly measured in different studies, including different muscle groups, contraction types and measurement instruments. Moreover, different genes are likely to contribute to different aspects of strength that may not be reflected across the different measurement types. All this means that for a particular gene or genotype of interest, the chances of replication across multiple studies are small. But when genes are found to be important across multiple, different strength measurements the likelihood the gene is important to muscle strength improves considerably.

**Angiotensin-converting enzyme (ACE).** *ACE* and its insertion/deletion polymorphism have been studied in relation to

skeletal muscle traits in a number of studies. Though some studies have reported significant associations with baseline muscle strength phenotypes,<sup>59–65</sup> there are inconsistent associations within those studies and several other investigations have not observed significant associations.<sup>66–72</sup> Thus, there is little evidence to suggest that *ACE* genotype is a strong contributor to inter-individual variation in skeletal muscle strength.

**Alpha actinin 3 (*ACTN3*).** The *ACTN3* gene has generated considerable attention following a number of cross-sectional investigations in elite athletes that pointed to a disadvantage for homozygote carriers of the R577X nonsense (X) allele in sprint and power-related activities.<sup>73–75</sup> When examining the breadth of studies in this area, most point to slightly lower muscle strength values in X/X vs R-allele carriers,<sup>76–78</sup> though not all studies support this conclusion.<sup>69,79–81</sup> For example, Vincent and colleagues<sup>76</sup> studied the R577X polymorphism in relation to isometric and isokinetic knee extensor strength in 90 young men and reported lower concentric peak torque at 300° s<sup>-1</sup> in X/X compared with R/R homozygotes. The authors also reported a lower proportion of type IIx muscle fibers in X/X vs R/R homozygotes. In contrast, Norman and colleagues<sup>80</sup> reported no significant associations with muscle power or torque-velocity relationships among *ACTN3* genotypes in a study of 120 moderately to well-trained men and women. They were also unable to confirm the difference in fiber-type proportion reported by Vincent and colleagues. Interestingly, in a longitudinal study of 1367 older adults (70–79 years), Delmonico *et al.*<sup>82</sup> reported greater losses of 400 m walk time performance over 5 years in male X/X vs R-allele carriers, whereas X/X women had a 35% greater risk of lower extremity physical limitation compared with R/R women. Judson *et al.*<sup>83</sup> recently reported great risk of falling in older females carrying at least one X allele. The general consensus among these studies is that *ACTN3* X/X carriers have modestly lower skeletal muscle strength and power in comparison with R-allele carriers, with the work of Delmonico *et al.*<sup>82</sup> and most recently Judson *et al.*<sup>83</sup> indicating potential clinical importance for the X/X genotype in older men and women.

**Ciliary neurotrophic factor (*CNTF*).** Several studies have examined genetic variation in the *CNTF* gene and/or its receptor *CNTFR*. A rare null allele in the *CNTF* gene has been associated with muscle strength,<sup>84,85</sup> but the frequency of the rare A/A genotype is so low that general public health significance is unclear even while it might be clinically important for those particular individuals. Multiple polymorphisms in the *CNTFR* gene have also been studied in relation to strength variables<sup>86,87</sup> but no consistent findings have been observed across studies. Overall, these findings indicate the potential for significant influences of *CNTF* and *CNTFR* gene variants on skeletal muscle strength, with the rare A/A genotype in *CNTF* appearing to have the most clinical relevance.

**Myostatin-related genes.** Myostatin emerged as an attractive target of gene-association studies and multiple polymorphisms were identified in the human gene (*MSTN*).<sup>88</sup> Some investigations have reported associations with skeletal muscle strength, but the sample sizes are very small owing in part to low allele frequencies of the common polymorphisms.<sup>89–91</sup> Because the common polymorphisms have rare allele frequencies, any public

health significance of *MSTN* genetic variation is unlikely, though it may be important for those individuals. Subsequently genes within the myostatin-signaling pathway have been examined, including the myostatin receptors (*ACVR1B* and *ACVR2B*) and follistatin, a myostatin inhibitor,<sup>55,92,93</sup> but again, the sample sizes of the genotype groups with significant findings were generally small, making the clinical relevance of these findings uncertain but generally not striking.

**Vitamin D receptor (*VDR*).** Vitamin D deficiency has been consistently associated with lower muscle strength<sup>94</sup> and has been discussed as a potential mechanism of sarcopenia.<sup>95</sup> The *VDR* gene has multiple polymorphisms that have been investigated but studies differ with regard to the specific polymorphisms or haplotypes examined, making comparisons difficult. *VDR* genetic variation has been associated with muscle strength variables in numerous studies,<sup>96–103</sup> though inconsistencies have been noted because of the examination of different variants. Studies having examined the BsmI locus are more mixed with regard to their findings and future studies need to incorporate the haplotype of BsmI and TaqI rather than looking at either site independently. The *VDR* FokI site is considered functional<sup>104,105</sup> and two studies reported higher strength in f/f compared with F/F carriers.<sup>99,101</sup> Thus, the *VDR* locus is a promising target that should be investigated more thoroughly in future studies.

### Genetic Variation and Skeletal Muscle Mass

This section examines genes that have been studied in relation to skeletal muscle mass measurements, again focusing on frequently studied genes examined in multiple studies and associated with baseline muscle mass values.

***ACE*.** The majority of papers examining the *ACE* insertion/deletion polymorphism have been focused on muscle strength rather than muscle mass phenotypes, though some studies have examined both.<sup>64,67,68,70</sup> Most have reported no association with muscle mass and it appears unlikely that *ACE* genotype contributes significantly to muscle mass phenotypes.

***ACTN3*.** Several studies have examined the potential for the *ACTN3* R577X polymorphism to explain variability in muscle strength measures and many of those same papers have examined muscle mass variables.<sup>76,78,80,82,106</sup> Of those studies, only Walsh *et al.*<sup>78</sup> and Zempo *et al.*<sup>106</sup> found evidence of an association between muscle size and the *ACTN3* null X allele, indicating at best a minor role for this polymorphism in explaining inter-individual variability in this trait.

**Androgen receptor (*AR*).** A few studies have examined the association between the *AR* CAG-repeat polymorphism and muscle mass variables with conflicting results, with both longer repeat length<sup>107,108</sup> and shorter repeat length<sup>109</sup> being correlated with greater fat-free mass (FFM). Walsh and colleagues<sup>108</sup> found significant genotype associations with FFM in men from two independent cohorts and similar results were found by another group.<sup>107</sup> Nielsen *et al.*<sup>109</sup> observed opposite results but in a cohort of young men, indicating the possibility of an age interaction. Recent data indicate that the CAG-repeat sequence in the *AR* gene modulates receptor transcriptional activity and affects muscle cell development in culture.<sup>110</sup> Additional work is required to clarify these findings.



**Myostatin-related genes.** Despite the strong physiological evidence behind myostatin as a candidate gene for muscle mass traits and the importance of rare mutations in the gene on muscle mass,<sup>111</sup> common genetic variation in the *MSTN* gene has not been associated with muscle mass.<sup>93,112</sup> Studies that have examined myostatin-related genes in relation to muscle mass phenotypes have produced some minor associations<sup>92,93</sup> but there is little compelling evidence that *MSTN* or myostatin-related genes are major contributors to skeletal muscle mass.

**TRHR.** As described above, Liu and colleagues<sup>56</sup> identified *TRHR* as a potential candidate gene for skeletal muscle mass from the first GWAS for this trait. The authors performed separate replication studies in three cohorts consisting of over 6000 total white and Chinese subjects and consistent significant associations with lean body mass were observed in those analyses. Though only a single paper, the multiple replications pointing to *TRHR* provide strength for this as a potentially important candidate gene for muscle mass variation.

**VDR.** Though *VDR* genetic variation has been studied extensively in relation to muscle strength, fewer studies have focused on skeletal muscle mass. Van Pottelbergh and colleagues<sup>113</sup> reported associations between TaqI/ApaI haplotypes and lean mass in 271 older men (>70 years), but not in a separate group of younger men from the same study. Roth *et al.*<sup>99</sup> reported significant associations with the *VDR* FokI polymorphism (f and F alleles) and leg FFM in 302 older Caucasian men, with concomitant differences in muscle strength. These results provide evidence for positive association and continued interest in this gene in relation to skeletal muscle traits.

### Genetic Variation and Sarcopenia

Though a number of studies have investigated specific genes and genetic variants in relation to skeletal muscle strength and mass phenotypes, only one study has specifically targeted sarcopenia *per se*. Roth and colleagues<sup>99</sup> analyzed the influence of *VDR* sequence variants on muscle strength and mass in a cohort of 302 older (58–93 years) Caucasian men with measures of FFM by dual-energy X-ray absorptiometry. *VDR* FokI genotype was significantly associated with different lean mass measures, with the F/F group demonstrating significantly lower mass than the F/f and f/f groups. In addition, the group categorized the men as normal or sarcopenic based on the definition of Baumgartner *et al.*,<sup>3</sup> which relies on a cutoff value based on appendicular FFM relative to body weight ( $\text{kg m}^{-2}$ ). Logistic regression revealed a twofold higher risk for sarcopenia in *VDR* FokI F/F homozygotes than carriers of the f allele. Quadriceps muscle strength was also lower in the F/F group compared with the F/f and f/f groups, but this association was eliminated when the analysis controlled for the differences in total body lean mass. Thus, *VDR* FokI genotype was significantly associated with lean mass and sarcopenia in this cohort of older Caucasian men, with concomitant differences in muscle strength. Additional work is needed in this area.

### Conclusions and Future Directions

Though high heritability values would indicate a strong genetic component, little progress has been made in identifying specific genetic contributors to skeletal muscle strength and mass

phenotypes relevant to sarcopenia. Although many genes have been tentatively examined (not all reviewed here), few have been positively associated with muscle-related traits across multiple cohorts and the findings are not always consistent within any replication analyses. No genes have been replicated for association with sarcopenia itself, though *VDR* genotypes have been associated with sarcopenia in one study and with muscle strength and mass phenotypes in multiple studies.

Of those genes that have been identified, their importance to skeletal muscle-trait variation is generally small. None of the genes described above have been shown to conclusively contribute >5% of the inter-individual variation to their respective traits, and most are on the order of 1–3%. In addition to typical polymorphisms, copy number variation (multiple copies of the same gene), gene–gene interactions (multiple genes coordinated in a pathway), complex gene–environment interactions and epigenetic factors also contribute to the genetic component of inter-individual variability,<sup>114</sup> and these more complex phenomena are just beginning to be studied in large-scale investigations. One possible approach to addressing this complexity is the calculation of genetic predisposition scores based on multiple genetic variants.<sup>115</sup> Finally, it is important to recognize that genetic factors will only be one of the several contributors to sarcopenia-related traits and additional environmental and developmental factors must not be ignored when addressing future prevention and treatment strategies.<sup>116,117</sup>

A consideration when examining the genetic aspects of skeletal muscle traits generally and sarcopenia in particular is that of a ‘threshold’ level for these traits below which physical function is impaired. Once strength falls below a threshold value specific to that individual, physical function is impacted. Though such a threshold would surely be different for each individual, we can expect clinically meaningful thresholds could be established across various physical characteristics, including body composition and strength. This threshold concept has been discussed by a number of groups.<sup>118–121</sup> Because genetic variation will tend to have subtle influences on skeletal muscle traits, that genetic influence will tend to push muscle trait values closer to or farther away from this threshold, altering an individual’s risk for impaired physical function. Identifying individuals with a genetic susceptibility to lower levels of skeletal muscle strength or mass (and who are closer to their likely threshold for physical limitation) will allow for earlier, targeted interventions to help prevent those losses. Similarly, it is important to recognize that genetic factors may have an impact on either the development of adult muscle mass and strength or the decline of these traits from their peak values in early adulthood. Different genetic influences can be envisioned for both of those traits, in effect differentiating the trajectories of muscle development from those of muscle loss. Finding these genes and developing the individualized interventions will take many years. Even if genes of only minor effect are identified that don’t lend themselves directly to genetic screening and personalized medicine, those genes will point to the potential physiological pathways that can be manipulated through more typical means and thereby add to our understanding of the underlying etiology of sarcopenia.<sup>122,123</sup>

### Conflict of Interest

The author declares no conflict of interest.

## References

1. Dutta C, Hadley EC. The significance of sarcopenia in old age. *J Gerontol Series A* 1995;**50A**:1–4.
2. Lindle RS, Metter EJ, Lynch NA, Fleg JL, Fozard JL, Tobin JD *et al*. Age and gender comparisons of muscle strength in 654 women and men aged 20–93 yr. *J Appl Physiol* 1997;**83**:1581–1587.
3. Baumgartner RN, Koehler KM, Gallagher D, Romero L, Heymsfield SB, Ross RR *et al*. Epidemiology of sarcopenia among the elderly in New Mexico. *Am J Epidemiol* 1998;**147**:755–763.
4. Janssen I, Heymsfield SB, Ross R. Low relative skeletal muscle mass (sarcopenia) in older persons is associated with functional impairment and physical disability. *J Am Geriatr Soc* 2002;**50**:889–896.
5. Lauretani F, Russo CR, Bandinelli S, Bartali B, Cavazzini C, Di Iorio A *et al*. Age-associated changes in skeletal muscles and their effect on mobility: an operational diagnosis of sarcopenia. *J Appl Physiol* 2003;**95**:1851–1860.
6. Newman AB, Kupelian V, Visser M, Simonsick E, Goodpaster BH, Nevitt M *et al*. Sarcopenia: alternative definitions and associations with lower extremity function. *J Am Geriatr Soc* 2003;**51**:1602–1609.
7. Visser M, Deeg DJH, Lips P, Harris TB, Bouter LM. Skeletal muscle mass and muscle strength in relation to lower-extremity performance in older men and women. *J Am Geriatr Soc* 2000;**48**:381–386.
8. Kwon IS, Oldaker S, Schrage MA, Talbot LA, Fozard JL, Metter EJ. Relationship between muscle strength and the time taken to complete a standardized walk-turn-walk test. *J Gerontol Biol Sci* 2001;**56A**:B398–B404.
9. Rantanen T, Guralnik JM, Izmirlian G, Williamson JD, Simonsick EM, Ferrucci L *et al*. Association of muscle strength with maximum walking speed in disabled older women. *Am J Phys Med Rehabil* 1998;**77**:299–305.
10. Buchner DM, deLateur B. The importance of skeletal muscle strength to physical function in older adults. *Ann Behav Med* 1991;**13**:91–98.
11. Nybo H, Gaist D, Jeune B, McGue M, Vaupel JW, Christensen K. Functional status and self-rated health in 2,262 nonagenarians: the Danish 1905 Cohort Survey. *J Am Geriatr Soc* 2001;**49**:601–609.
12. Rantanen T, Volpato S, Ferrucci L, Heikkinen E, Fried LP, Guralnik JM. Handgrip strength and cause-specific and total mortality in older disabled women: exploring the mechanism. *J Am Geriatr Soc* 2003;**51**:636–641.
13. Goodpaster BH, Park SW, Harris TB, Kritchevsky SB, Nevitt M, Schwartz AV *et al*. The loss of skeletal muscle strength, mass, and quality in older adults: the health, aging and body composition study. *J Gerontol A Biol Sci Med Sci* 2006;**61**:1059–1064.
14. Metter EJ, Talbot LA, Schrage MA, Conwit R. Skeletal muscle strength as a predictor of all-cause mortality in healthy men. *J Gerontol Biol Sci* 2002;**57**:B359–B365.
15. Rantanen T, Harris T, Leveille SG, Visser M, Foley D, Masaki K *et al*. Muscle strength and body mass index as long-term predictors of mortality in initially healthy men. *J Gerontol Med Sci* 2000;**55A**:M168–M173.
16. Newman AB, Kupelian V, Visser M, Simonsick EM, Goodpaster BH, Kritchevsky SB *et al*. Strength, but not muscle mass, is associated with mortality in the health, aging and body composition study cohort. *J Gerontol Med Sci* 2006;**61A**:72–77.
17. Janssen I, Shepard DS, Katzmarzyk PT, Roubenoff R. The healthcare costs of sarcopenia in the United States. *J Am Geriatr Soc* 2004;**52**:80–85.
18. Melton LJ, Khosla S, Crowley CS, O'Connor MK, O'Fallon WM, Riggs BL. Epidemiology of sarcopenia. *J Am Geriatr Soc* 2000;**48**:625–630.
19. Tanko LB, Movsesyan L, Mouritzen U, Christiansen C, Svendsen OL. Appendicular lean tissue mass and the prevalence of sarcopenia among healthy women. *Metabolism* 2002;**51**:69–74.
20. Sowers MR, Crutchfield M, Richards K, Wilkin MK, Furniss A, Jannausch M *et al*. Sarcopenia is related to physical functioning and leg strength in middle-aged women. *J Gerontol Biol Sci* 2005;**60A**:486–490.
21. Hurley BF, Hanson ED, Sheaff AK. Strength training as a countermeasure to aging muscle and chronic disease. *Sports Med* 2011;**41**:289–306.
22. Peterson MD, Sen A, Gordon PM. Influence of resistance exercise on lean body mass in aging adults: a meta-analysis. *Med Sci Sports Exerc* 2011;**43**:249–258.
23. Thomis MAI, Beunen GP, Maes HH, Blimkie CJ, Van Leemputte M, Claessens AL *et al*. Strength training: importance of genetic factors. *Med Sci Sports Exerc* 1998;**30**:724–731.
24. Bray MS, Hagberg JM, Perusse L, Rankinen T, Roth SM, Wolfarth B *et al*. The human gene map for performance and health-related fitness phenotypes: the 2006–2007 update. *Med Sci Sports Exerc* 2009;**41**:35–73.
25. Tan LJ, Liu SL, Lei SF, Papanicolaou CJ, Deng HW. Molecular genetic studies of gene identification for sarcopenia. *Hum Genet* 2012;**131**:1–31.
26. Montoye HJ, Metzner HL, Keller JB. Familial aggregation of strength and heart rate response to exercise. *Hum Biol* 1975;**47**:17–36.
27. Venerando A, Milani-Comparetti M. Twin studies in sport and physical performance. *Acta Genet Med Gemellol (Roma)* 1970;**19**:80–82.
28. Kovar R. Letter: motor performances in twins. *Acta Genet Med Gemellol (Roma)* 1975;**24**:174.
29. Reed T, Fabsitz RR, Selby JV, Carmelli D. Genetic influences and grip strength norms in the NHLBI twin study males aged 59–69. *Ann Hum Biol* 1991;**18**:425–432.
30. Thomis MAI, Van Leemputte M, Maes HH, Blimkie CJ, Claessens AL, Marchal G *et al*. Multivariate genetic analysis of maximal isometric muscle force at different elbow angles. *J Appl Physiol* 1997;**82**:959–967.
31. Thomis MAI, Beunen GP, Van Leemputte M, Maes HH, Blimkie CJ, Claessens AL *et al*. Inheritance of static and dynamic arm strength and some of its determinants. *Acta Physiol Scand* 1998;**163**:59–71.
32. Huygens W, Thomis MA, Peeters MW, Vlietinck RF, Beunen GP. Determinants and upper-limit heritabilities of skeletal muscle mass and strength. *Can J Appl Physiol* 2004;**29**:186–200.
33. Karlsson J, Komi PV, Viitasalo JHT. Muscle strength and muscle characteristics in monozygous and dizygous twins. *Acta Physiol Scand* 1979;**106**:319–325.
34. Bouchard C, Savard R, Despres JP, Tremblay A, LeBlanc C. Body composition in adopted and biological siblings. *Hum Biol* 1985;**57**:61–75.
35. Forbes GB, Sauer EP, Weitkamp LR. Lean body mass in twins. *Metabolism* 1995;**44**:1442–1446.
36. Seeman E, Hopper JL, Young NR, Formica C, Goss P, Tsalamandris C. Do genetic factors explain associations between muscle strength, lean mass, and bone density? A twin study. *Am J Physiol* 1996;**270**:E320–E327.
37. Arden NK, Spector TD. Genetic influences on muscle strength, lean body mass, and bone mineral density: a twin study. *J Bone Mineral Res* 1997;**12**:2076–2081.
38. Loos R, Thomis MAI, Maes HH, Beunen GP, Claessens AL, Derom C *et al*. Gender-specific regional changes in genetic structure of muscularity in early adolescence. *J Appl Physiol* 1997;**82**:1802–1810.
39. Frederiksen H, Gaist D, Petersen HC, Hjelmborg J, McGue M, Vaupel JW *et al*. Hand grip strength: a phenotype suitable for identifying genetic variants affecting mid- and late-life physical functioning. *Genet Epidemiol* 2002;**23**:110–122.
40. Tiainen K, Sipilä S, Alen M, Heikkinen E, Kaprio J, Koskenvuo M *et al*. Heritability of maximal isometric muscle strength in older female twins. *J Appl Physiol* 2004;**96**:173–180.
41. Zhai G, Stankovich J, Ding C, Scott F, Cicuttini F, Jones G. The genetic contribution to muscle strength, knee pain, cartilage volume, bone size, and radiographic osteoarthritis: a sibpair study. *Arthritis Rheum* 2004;**50**:805–810.
42. Tiainen K, Sipilä S, Kauppinen M, Kaprio J, Rantanen T. Genetic and environmental effects on isometric muscle strength and leg extensor power followed up for three years among older female twins. *J Appl Physiol* 2009;**106**:1604–1610.
43. Tiainen K, Sipilä S, Alen M, Heikkinen E, Kaprio J, Koskenvuo M *et al*. Shared genetic and environmental effects on strength and power in older female twins. *Med Sci Sports Exerc* 2005;**37**:72–78.
44. Carmelli D, Kelly-Hayes M, Wolf PA, Swan GE, Jack LM, Reed T *et al*. The contribution of genetic influences to measures of lower-extremity function in older male twins. *J Gerontol Biol Sci* 2000;**55A**:B49–B53.
45. Zhai G, Ding C, Stankovich J, Cicuttini F, Jones G. The genetic contribution to longitudinal changes in knee structure and muscle strength: a sibpair study. *Arthritis Rheum* 2005;**52**:2830–2834.
46. Carmelli D, Reed T. Stability and change in genetic and environmental influences on hand-grip strength in older male twins. *J Appl Physiol* 2000;**89**:1879–1883.
47. Chagnon YC, Borecki I, Perusse L, Roy S, Lacaille M, Chagnon M *et al*. Genome-wide search for genes related to the fat-free body mass in the Quebec Family Study. *Metabolism* 2000;**49**:203–207.
48. Livshits G, Kato BS, Wilson SG, Spector TD. Linkage of genes to total lean body mass in normal women. *J Clin Endocrinol Metab* 2007;**92**:3171–3176.
49. Karasik D, Zhou Y, Cupples LA, Hannan MT, Kiel DP, Demissie S. Bivariate genome-wide linkage analysis of femoral bone traits and leg lean mass: Framingham study. *J Bone Miner Res* 2009;**24**:710–718.
50. De Mars G, Windelinckx A, Huygens W, Peeters MW, Beunen GP, Aerssens J *et al*. Genome-wide linkage scan for contraction velocity characteristics of knee musculature in the Leuven Genes for Muscular Strength Study. *Physiol Genomics* 2008;**35**:36–44.
51. De Mars G, Windelinckx A, Huygens W, Peeters MW, Beunen GP, Aerssens J *et al*. Genome-wide linkage scan for maximum and length-dependent knee muscle strength in young men: significant evidence for linkage at chromosome 14q24.3. *J Med Genet* 2008;**45**:275–283.
52. Sun G, Gagnon J, Chagnon YC, Perusse L, Despres JP, Leon AS *et al*. Association and linkage between an insulin-like growth factor-1 gene polymorphism and fat free mass in the HERITAGE Family Study. *Int J Obesity* 1999;**23**:929–935.
53. Huygens W, Thomis MA, Peeters MW, Aerssens J, Janssen R, Vlietinck RF *et al*. Linkage of myostatin pathway genes with knee strength in men. *Physiol Genomics* 2004;**17**:264–270.
54. Huygens W, Thomis MA, Peeters MW, Aerssens J, Vlietinck R, Beunen GP. Quantitative trait loci for human muscle strength: linkage analysis of myostatin pathway genes. *Physiol Genomics* 2005;**22**:390–397.
55. Windelinckx A, De Mars G, Huygens W, Peeters MW, Vincent B, Wijmenga C *et al*. Comprehensive fine mapping of chr12q12-14 and follow-up replication identify activin receptor 1B (ACVR1B) as a muscle strength gene. *Eur J Hum Genet* 2011;**19**:208–215.
56. Liu XG, Tan LJ, Lei SF, Liu YJ, Shen H, Wang L *et al*. Genome-wide association and replication studies identified TRHR as an important gene for lean body mass. *Am J Hum Genet* 2009;**84**:418–423.
57. Sun L, Tan LJ, Lei SF, Chen XD, Li X, Pan R *et al*. Bivariate genome-wide association analyses of femoral neck bone geometry and appendicular lean mass. *PLoS One* 2011;**6**:e27325.
58. Hai R, Pei YF, Shen H, Zhang L, Liu XG, Lin Y *et al*. Genome-wide association study of copy number variation identified gremlin1 as a candidate gene for lean body mass. *J Hum Genet* 2012;**57**:33–37.
59. Woods D, Onambele G, Woledge R, Skelton D, Bruce S, Humphries SE *et al*. Angiotensin-I converting enzyme genotype-dependent benefit from hormone replacement therapy in isometric muscle strength and bone mineral density. *J Clin Endocrinol Metab* 2001;**86**:2200–2204.

60. Hopkinson NS, Nickol AH, Payne J, Hawe E, Man WD, Moxham J *et al.* Angiotensin converting enzyme genotype and strength in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2004;**170**:395–399.
61. Williams AG, Day SH, Folland JP, Gohlike P, Dhamrait S, Montgomery HE. Circulating angiotensin converting enzyme activity is correlated with muscle strength. *Med Sci Sports Exerc* 2005;**37**:944–948.
62. Moran CN, Vassilopoulos C, Tsiokanos A, Jamurtas AZ, Bailey ME, Montgomery HE *et al.* The associations of ACE polymorphisms with physical, physiological and skill parameters in adolescents. *Eur J Hum Genet* 2006;**14**:332–339.
63. Wagner H, Thaller S, Dahse R, Sust M. Biomechanical muscle properties and angiotensin-converting enzyme gene polymorphism: a model-based study. *Eur J Appl Physiol* 2006;**98**:507–515.
64. Charbonneau DE, Hanson ED, Ludlow AT, Delmonico MJ, Hurley BF, Roth SM. ACE genotype and the muscle hypertrophic and strength responses to strength training. *Med Sci Sports Exerc* 2008;**40**:677–683.
65. Yoshihara A, Tobina T, Yamaga T, Ayabe M, Yoshitake Y, Kimura Y *et al.* Physical Function Is Weakly Associated with Angiotensin-Converting Enzyme Gene I/D Polymorphism in Elderly Japanese Subjects. *Gerontology* 2009;**55**:387–392.
66. Folland J, Leach B, Little T, Hawker K, Myerson S, Montgomery H *et al.* Angiotensin-converting enzyme genotype affects the response of human skeletal muscle to functional overload. *Exp Physiol* 2000;**85**:575–579.
67. Thomis MA, Huygens W, Heuninckx S, Chagnon M, Maes HHM, Claessens AL *et al.* Exploration of myostatin polymorphisms and the angiotensin-converting enzyme insertion/deletion genotype in responses of human muscle to strength training. *Eur J Appl Physiol* 2004;**92**:267–274.
68. Pescatello LS, Kostek MA, Gordish-Dressman H, Thompson PD, Seip RL, Price TB *et al.* ACE ID genotype and the muscle strength and size response to unilateral resistance training. *Med Sci Sports Exerc* 2006;**38**:1074–1081.
69. McCauley T, Mastana SS, Hossack J, Macdonald M, Folland JP. Human angiotensin-converting enzyme I/D and alpha-actinin 3 R577X genotypes and muscle functional and contractile properties. *Exp Physiol* 2009;**94**:81–89.
70. Lima RM, Leite TK, Pereira RW, Rabelo HT, Roth SM, Oliveira RJ. ACE and ACTN3 genotypes in older women: muscular phenotypes. *Int J Sports Med* 2011;**32**:66–72.
71. Rodriguez-Romo G, Ruiz JR, Santiago C, Fiuza-Luces C, Gonzalez-Freire M, Gomez-Gallego F *et al.* Does the ACE I/D polymorphism, alone or in combination with the ACTN3 R577X polymorphism, influence muscle power phenotypes in young, non-athletic adults? *Eur J Appl Physiol* 2010;**110**:1099–1106.
72. Bustamante-Ara N, Santiago C, Verde Z, Yvert T, Gomez-Gallego F, Rodriguez-Romo G *et al.* ACE and ACTN3 genes and muscle phenotypes in nonagenarians. *Int J Sports Med* 2010;**31**:221–224.
73. Yang N, MacArthur DG, Gulbin JP, Hahn AG, Beggs AH, Eastale S *et al.* ACTN3 genotype is associated with human elite athletic performance. *Am J Hum Genet* 2003;**73**:627–631.
74. Niemi AK, Majamaa K. Mitochondrial DNA and ACTN3 genotypes in Finnish elite endurance and sprint athletes. *Eur J Hum Genet* 2005;**13**:965–969.
75. Roth SM, Walsh S, Liu D, Metter EJ, Ferrucci L, Hurley BF. The ACTN3 R577X nonsense allele is under-represented in elite-level strength athletes. *Eur J Hum Genet* 2008;**16**:391–394.
76. Vincent B, De Boek K, Ramaekers M, Van den Eede E, Van Leemputte M, Hespel P *et al.* ACTN3 (R577X) genotype is associated with fiber type distribution. *Physiol Genomics* 2007;**32**:58–63.
77. Clarkson PM, Devaney JM, Gordish-Dressman H, Thompson PD, Hubal MJ, Urso M *et al.* ACTN3 genotype is associated with increases in muscle strength in response to resistance training in women. *J Appl Physiol* 2005;**99**:154–163.
78. Walsh S, Liu D, Metter EJ, Ferrucci L, Roth SM. ACTN3 genotype is associated with muscle phenotypes in women across the adult age span. *J Appl Physiol* 2008;**105**:1486–1491.
79. Delmonico MJ, Kostek MC, Doldo NA, Hand BD, Walsh S, Conway JM *et al.* Alpha-actinin-3 (ACTN3) R577X polymorphism influences knee extensor peak power response to strength training in older men and women. *J Gerontol A Biol Sci Med Sci* 2007;**62**:206–212.
80. Norman B, Esbjornsson M, Rundqvist H, Osterlund T, von Walden F, Tesch PA. Strength, power, fiber types, and mRNA expression in trained men and women with different ACTN3 R577X genotypes. *J Appl Physiol* 2009;**106**:959–965.
81. Hanson ED, Ludlow AT, Sheaff AK, Park J, Roth SM. ACTN3 genotype does not influence muscle power. *Int J Sports Med* 2010;**31**:834–838.
82. Delmonico MJ, Zmuda JM, Taylor BC, Cauley JA, Harris TB, Manini TM *et al.* Association of the ACTN3 genotype and physical functioning with age in older adults. *J Gerontol A Biol Sci Med Sci* 2008;**63**:1227–1234.
83. Judson RN, Wackerhage H, Hughes A, Mavroidi A, Barr RJ, Macdonald HM *et al.* The functional ACTN3 577X variant increases the risk of falling in older females: results from two large independent cohort studies. *J Gerontol A Biol Sci Med Sci* 2011;**66**:130–135.
84. Roth SM, Schragger MA, Ferrell RE, Reichman SE, Metter EJ, Lynch NA *et al.* CNTF genotype is associated with muscular strength and quality in humans across the adult age span. *J Appl Physiol* 2001;**90**:1205–1210.
85. Arking DE, Fallin DM, Fried LP, Li T, Beamer BA, Xue QL *et al.* Variation in the ciliary neurotrophic factor gene and muscle strength in older Caucasian women. *J Am Geriatr Soc* 2006;**54**:823–826.
86. Roth SM, Metter EJ, Lee MR, Hurley BF, Ferrell RE. C174T polymorphism in the CNTF receptor gene is associated with fat-free mass in men and women. *J Appl Physiol* 2003;**95**:1425–1430.
87. De Mars G, Windelinckx A, Beunen G, Delecluse C, Lefevre J, Thomis MA. Polymorphisms in the CNTF and CNTF receptor genes are associated with muscle strength in men and women. *J Appl Physiol* 2007;**102**:1824–1831.
88. Ferrell RE, Conte V, Lawrence EC, Roth SM, Hagberg JM, Hurley BF. Frequent sequence variation in the human myostatin (GDF8) gene as a marker for analysis of muscle-related phenotypes. *Genomics* 1999;**62**:203–207.
89. Seibert MJ, Xue Q-L, Fried LP, Walston JD. Polymorphic variation in the human myostatin (GDF-8) gene and association with strength measures in the Women's Health and Aging Study II cohort. *J Am Geriatr Soc* 2001;**49**:1093–1096.
90. Corsi AM, Ferrucci L, Gozzini A, Tanini A, Brandi ML. Myostatin polymorphisms and age-related sarcopenia in the Italian population. *J Am Geriatr Soc* 2002;**50**:1463.
91. Santiago C, Ruiz JR, Rodriguez-Romo G, Fiuza-Luces C, Yvert T, Gonzalez-Freire M *et al.* The K153R polymorphism in the myostatin gene and muscle power phenotypes in young, non-athletic men. *PLoS One* 2011;**6**:e16323.
92. Walsh S, Metter EJ, Ferrucci L, Roth SM. Activin-type II receptor B (ACVR2B) and follistatin haplotype associations with muscle mass and strength in humans. *J Appl Physiol* 2007;**102**:2142–2148.
93. Kostek MA, Angelopoulos TJ, Clarkson PM, Gordon PM, Moyna NM, Visich PS *et al.* Myostatin and follistatin polymorphisms interact with muscle phenotypes and ethnicity. *Med Sci Sports Exerc* 2009;**41**:1063–1071.
94. Ceglia L. Vitamin D and skeletal muscle tissue and function. *Mol Aspects Med* 2008;**29**:407–414.
95. Montero-Odasso M, Duque G. Vitamin D in the aging musculoskeletal system: an authentic strength preserving hormone. *Mol Aspects Med* 2005;**26**:203–219.
96. Geusens P, Vandevyver C, VanHoof J, Cassiman J, Boonen S, Rous J. Quadriceps and grip strength are related to vitamin D receptor genotype in elderly nonobese women. *J Bone Mineral Res* 1997;**12**:2082–2088.
97. Vandevyver C, VanHoof J, Declerck K, Stinissen P, Vandervorst C, Michiels L *et al.* Lack of association between estrogen receptor genotypes and bone mineral density, fracture history, or muscle strength in elderly women. *J Bone Mineral Res* 1999;**14**:1576–1582.
98. Grundberg E, Brandstrom H, Ribom EL, Ljunggren O, Mallmin H, Kindmark A. Genetic variation in the human vitamin D receptor is associated with muscle strength, fat mass and body weight in Swedish women. *Eur J Endocrinol* 2004;**150**:323–328.
99. Roth SM, Zmuda JM, Cauley JA, Shea PR, Ferrell RE. Vitamin D receptor genotype is associated with fat-free mass and sarcopenia in elderly men. *J Gerontol Biol Sci* 2004;**59A**:10–15.
100. Wang P, Ma LH, Wang HY, Zhang W, Tian Q, Cao DN *et al.* Association between polymorphisms of vitamin D receptor gene Apal, Bsm1 and Taq1 and muscular strength in young Chinese women. *Int J Sports Med* 2006;**27**:182–186.
101. Windelinckx A, De Mars G, Beunen G, Aerssens J, Delecluse C, Lefevre J *et al.* Polymorphisms in the vitamin D receptor gene are associated with muscle strength in men and women. *Osteoporos Int* 2007;**18**:1235–1242.
102. Hopkinson NS, Li KW, Kehoe A, Humphries SE, Roughton M, Moxham J *et al.* Vitamin D receptor genotypes influence quadriceps strength in chronic obstructive pulmonary disease. *Am J Clin Nutr* 2008;**87**:385–390.
103. Bahat G, Saka B, Erten N, Ozbek U, Coskunpinar E, Yildiz S *et al.* Bsm1 polymorphism in the vitamin D receptor gene is associated with leg extensor muscle strength in elderly men. *Aging Clin Exp Res* 2010;**22**:198–205.
104. Arai H, Miyamoto K, Taketani Y, Yamamoto H, Iemori Y, Morita K *et al.* A vitamin D receptor gene polymorphism in the translation initiation codon: effect on protein activity and relation to bone mineral density in Japanese women. *J Bone Min Res* 1997;**12**:915–921.
105. Jurutka PW, Remus LS, Whitfield GK, Thompson PD, SHsieh J-c, Zitzer H *et al.* The polymorphic N terminus in human vitamin D receptor isoforms influences transcriptional activity by modulating interaction with transcription factor IIB. *Mol Endocrinol* 2000;**14**:401–420.
106. Zempo H, Tanabe K, Murakami H, Iemitsu M, Maeda S, Kuno S. ACTN3 polymorphism affects thigh muscle area. *Int J Sports Med* 2010;**31**:138–142.
107. Campbell BC, Gray PB, Eisenberg DT, Ellison P, Sorenson MD. Androgen receptor CAG repeats and body composition among Atrial men. *Int J Androl* 2009;**32**:140–148.
108. Walsh S, Zmuda JM, Cauley JA, Shea PR, Metter EJ, Hurley BF *et al.* Androgen receptor CAG repeat polymorphism is associated with fat-free mass in men. *J Appl Physiol* 2005;**98**:132–137.
109. Nielsen TL, Hagen C, Wraae K, Bathum L, Larsen R, Brixen K *et al.* The impact of the CAG repeat polymorphism of the androgen receptor gene on muscle and adipose tissues in 20–29-year-old Danish men: Odense Androgen Study. *Eur J Endocrinol* 2010;**162**:795–804.
110. Sheppard RL, Spangenberg EE, Chin ER, Roth SM. Androgen receptor polyglutamine repeat length affects receptor activity and C2C12 cell development. *Physiol Genomics* 2011;**43**:1135–1143.
111. Schuelke M, Wagner KR, Stolz LE, Hubner C, Riebel T, Komen W *et al.* Myostatin mutation associated with gross muscle hypertrophy in a child. *N Engl J Med* 2004;**350**:2682–2688.
112. Ivey FM, Roth SM, Ferrell RE, Tracy BL, Lemmer JT, Hurlbut DE *et al.* Effects of age, gender, and myostatin genotype on the hypertrophic response to heavy resistance strength training. *J Gerontol A Biol Sci Med Sci* 2000;**55**:M641–M6M8.



113. Van Pottelbergh I, Goemaere S, De Bacquer D, De Paepe A, Kaufman JM. Vitamin D receptor gene allelic variants, bone density, and bone turnover in community-dwelling men. *Bone* 2002;**31**:631–637.
114. Altshuler D, Daly MJ, Lander ES. Genetic mapping in human disease. *Science* 2008;**322**: 881–888.
115. Thanassoulis G, Peloso GM, Pencina MJ, Hoffmann U, Fox CS, Cupples LA *et al*. A genetic risk score is associated with incident cardiovascular disease and coronary artery calcium: The Framingham Heart Study. *Circ Cardiovasc Genet* 2012;**5**:113–121.
116. Rolland Y, Dupuy C, Abellan van Kan G, Gillette S, Vellas B. Treatment strategies for sarcopenia and frailty. *Med Clin North Am* 2011;**95**:427–438, ix.
117. Burton LA, Sumukadas D. Optimal management of sarcopenia. *Clin Interv Aging* 2010;**5**: 217–228.
118. Ferrucci L, Guralnik JM, Buchner D, Kasper J, Lamb SE, Simonsick EM *et al*. Departures from linearity in the relationship between measures of muscular strength and physical performance of the lower extremities: the Women's Health and Aging Study. *J Gerontol A Biol Sci Med Sci* 1997;**52**:M275–M285.
119. Walston J, Fried LP. Frailty and the older man. *Med Clin North Amer* 1999;**83**:1173–1194.
120. Visser M, Newman AB, Nevitt MC, Kritchevsky SB, Stamm EB, Goodpaster BH *et al*. Reexamining the sarcopenia hypothesis. Muscle mass versus muscle strength. Health, Aging, and Body Composition Study Research Group. *Ann NY Acad Sci* 2000;**904**: 456–461.
121. McNeil CJ, Doherty TJ, Stashuk DW, Rice CL. Motor unit number estimates in the tibialis anterior muscle of young, old, and very old men. *Muscle Nerve* 2005;**31**: 461–467.
122. Khoury MJ, Davis R, Gwinn M, Lindegren ML, Yoon P. Do we need genomic research for the prevention of common diseases with environmental causes? *Am J Epidemiol* 2005;**161**:799–805.
123. Burke W. Genomics as a probe for disease biology. *N Engl J Med* 2003;**349**:969–974.